

Demographic and Laboratory Parameters among Covid-19 Outpatients Versus and Hospitalized Patients

GHUROOB DALIL DHUMAD¹, HAIDER SABAH KADHIM², HAIDER AHMAD SHAMRAN³, HATEM DHEYAB ABED⁴¹Faculty of Medicine, Microbiology Department, Al-nahrain University, Baghdad, Iraq, PhD²Faculty of Medicine, Microbiology Department, Al-nahrain University, Baghdad, Iraq, PhD³Faculty of Medicine, Microbiology Department, Al-nahrain University, Baghdad, Iraq, PhD⁴L-Kindy Teaching Hospital, Baghdad, Iraq, Arab board in internal medicineCorresponding author: Ghuroob Dalil Dhumad, Email: dghuroob@gmail.com

ABSTRACT

The present study addresses the impact of age and gender on poor diagnosis COVID-19 disease and monitoring the levels of the serum biomarkers: LDH, D-dimer, CRP, TGF-B, IL-6 among COVID-19 patients. The study included 100 COVID-19 patients, recruited from hospitals in Baghdad governorate from February 2020 to May 2020 of both genders with media range of age of 38-59 years. Data revealed no significant impact of gender on COVID-19 disease outcome. Whilst age of participants was significantly higher ($P < 0.001$) in needed ICU/admission died patients (59.0 ± 12.67 years) compared to that of well-discharged patients (38.83 ± 11.74 years). Age had a positive significant correlation with D-dimer ($r = 0.276$, $p = 0.006$), LDH ($r = 0.318$, $p = 0.003$), ferritin ($r = 0.307$, $p = 0.005$), CRP ($r = 0.470$, $p < 0.001$), and WBC count ($r = 0.410$, $p < 0.001$). D-dimer, LDH, ferritin, and CRP levels were significantly higher ($P < 0.001$) in needed ICU/admission died patients' group compared to those recorded in well-discharged patients' group. The sensitivity and specificity of the tests at $P < 0.001$ at cut off value of D-dimer = 0.56 mg/ml, 285.7 U/L, 387.97 ng/ml, and 36.3 mg/L were (81 and 94%), (71 and 76%), (86 and 84%), and (91 and 84%), for D-dimer, LDH, ferritin, and CRP, respectively.

Keywords: COVID-19; age, gender; D-dimer; LDH; CRP; ferritin; IL-6; TGF-B

INTRODUCTION

Acute respiratory distress syndrome (ARDS) is the leading cause of death in COVID-19 (1). The actual reason for this being a prevalent immunopathological event for SARS-CoV-2, SARS-CoV, and MERS-CoV infections is unknown, however it is most likely due to a cytokine storm (1-4). Differentially expressed cytokines are produced at sites of tissue inflammation and released into the blood by a variety of cell types, including macrophages, lymphocytes, endothelial cells, epithelial cells, and fibroblasts, and are involved in the pathophysiology of COVID-19 (2). IL-6 is a significant pro-inflammatory mediator (3) with a chief role in the host's defense against contagions and tissue destruction in SARS-CoV-2 infection. Significant beneficial impacts of IL-6 blockade therapy using a humanized anti-IL-6 receptor antibody, tocilizumab was recently reported in cases with cytokine release syndrome (4).

Transforming growth factor beta 1 (TGF- β 1), a member of the transforming growth factor beta superfamily of cytokines, [5] is a soluble cytokine secreted mostly by inflammatory cells and a well-known regulator of immune responses. As a consequence of TGF- β activation pathways the triggered inflammation, apoptosis, and fibrosis, would lead to harshly destructive influences in the lungs and other tissues [5, 6].

In vitro study conducted by β , Boumaza et al. revealed the infection of monocytes and macrophages by SARS-CoV-2 result in TGF- β secretion [7]. Only one reports stated the increased TGF- β 1 levels in two different subsets of CD4⁺ immune cells of the COVID-19 patient [8].

SARS-CoV-2 is considered a dual disease: respiratory disease and a hematologic disease. Blood clots exist in COVID-19 patients and involve in deep venous thrombosis in lower extremities [9]. TGF- β is known to elicit the production of Factor XII (FXII), stands at the onset of the coagulation signaling of the intrinsic cascade [10]. Thrombin mediates the cleavage of fibrinogen to fibrin and results in mature TGF- β 1, creating a positive feedback loop [11].

As an inflammatory consequence induced by bacterial/viral infection or tissue destruction, the liver normally synthesizes the C-reactive protein (CRP) within 6-10 hrs of any tissue damage. Since COVID-19 infection is a status with hyper inflammatory response with a pathological dysfunction of innate host defense mechanisms accompanied by multiple organ failure and cytokines storm [12]. Hence, the CRP level during COVID-19 infection is a good indicative about the acuteness status of the illness and is a strong prognostic marker for COVID-19 poor outcome [13].

Lactate dehydrogenase (LDH), an intracellular enzyme found in all tissue, does catalyze the conversion of pyruvate to lactate and NADH and NAD⁺. High serum levels of LDH in any individual is an indicative of cytoplasmic cell's membrane damage induced by viral infection [14,15]. Elevated levels of LDH in COVID-19 patients are expected as long as the LDH is already exists in lung tissue. Additionally, intrathrombotic microangiopathy, which is correlated with renal failure and cardiac damage, LDH levels are high [16,17].

D-dimer, a plasmin cleavage product, is made up of two adjacent fibrin 'D' domains and released as a single molecule [18]. In SARS-CoV correlated coagulopathy, the high D-dimer levels might result from upregulated urokinase-type plasminogen activator [19]. High levels of D-dimer ($>1 \mu\text{g/mL}$) were reportedly to play a crucial role in poor prognosis in COVID-19 patients [20,21].

Ferritin levels on admission in COVID-19 patients were 1.5 to 5.3 times higher in patients with severe disease compared to those with less-severe disease [22,23].

The aim of the current study is monitoring the levels of some serum biomarkers like LDH, D-dimer, IL-6, LDH, and TGF-B in COVID-19 patients in Iraq.

MATERIALS AND METHODS

Study cohort: The study cohort encompassed 100 Iraqi COVID-19 patients (21 to 81 years, both genders), recruited from hospitals in Baghdad governorate; Ibn-Al Qiph hospital and Al-Kindi Teaching hospital from February 2020 to May 2020. All participants signed the ethical consent settled by the Institutional Review Board (IRB) of Al-Nahrain University.

Blood sampling: Around three ml of venous blood were collected from each patient in a sterile plane tube. After 15 min to allow clotting, the blood sample was centrifuged at 4,000 rpm for 15 min. The resultant serum was kept at -20°C until being processed.

Quantification of IL-6, TGF- β 1, D-dimer, LDH, and Ferritin serum level: The levels of Interleukin-6 (IL-6) and TGF- β 1 in serum samples were estimated using Sandwich-ELISA method using Horseradish Peroxidase (HRP)-conjugated antibody specific for IL-6 and TGF- β 1. ELISA was performed in Microelisa strip plate according to the instructions of the manufacturer.

The level of D-Dimer in serum samples was estimated using antigen-antibody reaction using anti-human D-Dimer antibodies coated on the latex particles according to a previously reported procedure (Poudel et al., 2021). LDH, ferritin, and CRP were

measured in fully automated analyzer called Beckman Coulter Analyzers,

Statistical analysis: SPSS software version 25.0 was used to analyse all of the data (SPSS, Chicago). The normality of continuous data was tested (Shapiro Wilk test). The mean and standard deviation of normally distributed data were calculated and examined using the Student t-test. The Mann Whitney U test was used to assess data having non-normal distributions, which were reported as median and range. The Chi-square test was used to examine categorical variables that were expressed as numbers and percentages. The numerous markers were evaluated using a receiver operating characteristic curve (ROC) to predict the COVID-19 severity. The possible link of several indicators with age and duration severity of COVID-19 was investigated using Pearson's correlation test and binary logistic regression to calculate ratios (OR) and their associated 95 percent confidence intervals (CI). A statistically significant difference was defined as a p-value less than 0.05.

RESULTS

Correlation of Demographic Characteristics with COVID-19 Outcome: The mean age of the required ICU/died patients (59.0±12.67 years) was significantly higher (P<0.001) than that of discharged patients (38.83±11.74 years)(Table 3-1). Conversely, no significant difference (P=0.217) was evidenced among the two groups of patients regarding the gender.

Table 1: Demographic characteristics of COVID-19 cohort participants (n=100)

Variable	Discharged well (n=77)	Need ICU admission/died (n=23)	p-value
Age, years Mean±SD	38.83±11.74	59.0±12.67	<0.001†
Gender			0.217 ‡
Male	46(59.74%)	17(73.91%)	
Female	31(40.26%)	6(26.09%)	

† Student t-test

‡ Chi square /exact Fisher tests

Correlation of Age and Duration with Different Serum Biomarkers: Spearman's correlation test was used to explore the possible correlation of age and duration with different biomarkers. Age had a positive significant correlation with D-dimer (r= 0.276, p= 0.006), LDH (r= 0.318, p= 0.003), ferritin (r= 0.307, p= 0.005), CRP (r= 0.470 p<0.001), and WBC count (r= 0.410, p<0.001) (Table 3-2). Transforming growth factor- β showed a negative significant correlation with viral load (r= -0.262, p= 0.015). However, disease duration showed a positive significant correlation with D-dimer (r= 0.319, p=0.002), LDH (0.265, p= 0.025), ferritin (r= 0.283, p= 0.008), and CRP (r= 0.349, p<0.001) (Table 3-2, Figures 3.1,3.2,3.3).

Table 2: Spearman's correlation of age and duration with different serum biomarkers

Variable	Age		Duration	
	r	p-value	r	p-value
D-dimer	0.276	0.006	0.319	0.002
LDH	0.318	0.003	0.265	0.025
Ferritin	0.307	0.005	0.283	0.008
CRP	0.470	<0.001	0.349	<0.001
IL-6	0.022	0.828	-0.016	0.880
TGF-β	0.145	0.180	-0.054	0.622
WBC	0.410	<0.001	0.148	0.215

Association of serum Biomarkers with Patients' Outcome: Data regarding laboratory parameters were found to be non-normally distributed. Accordingly, Mann Whitney U-test were used to compare these parameters between discharged well and required ICU/died patients. Only, four serum biomarkers (D-dimer, LDH, CRP, and ferritin) showed a significant association (P<0.001) with the patient's outcome (Table 3-3). (???)_The above-

mentioned four serum biomarkers exhibited higher levels than the normal levels in both patients' groups. The levels of these serum biomarkers were significantly higher (P<0.001) in the required ICU/died patients than those observed in the well-discharged patients' group. No significant difference was evidenced regarding IL-6 and TGF-B among the two patients' groups.

Table 3: Levels of serum biomarkers in COVID-19 patients.

Variables	Discharged well (n=77)	Need ICU admission/died(n=23)	p-value‡
D-dimer, ng/ml Median Range	0.2 0.02-1.56	1.55 0.18-8.44	<0.001
CRP, mg/L Mean±SD Range	18.2 0-119.65	58.0 7.44-17.4	<0.001
LDH, U/L Mean±SD Range	211.35 110-1808	601.0 117.0-1850	0.001
Ferritin, ng/ml Median Range	147.3 4.71-1000	706 227.8-1000	<0.001
IL-6, pg/ml Median Range	12.31 0.44-244.13	11.65 4.61-234.89	0.701
TGF-β, pg/ml Median Range	31.22 0.37-243.66	6.45 2.11-78.6	0.182

‡ Mann Whitney U test, P value significant at <0.05.

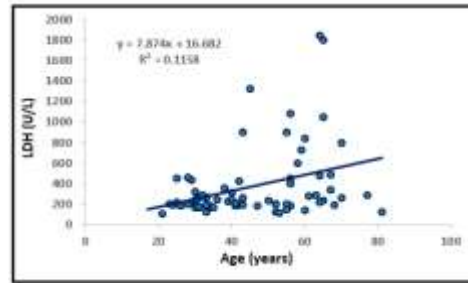


Figure 1: Scatter plot and regression line between age and LDH

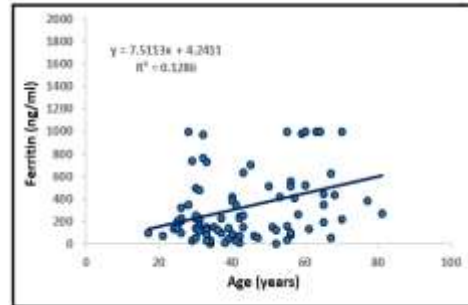


Figure 2: Scatter plot and regression line between age and ferritin

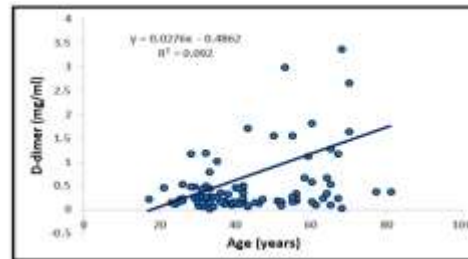


Figure 3: Scatter plot and regression line between age and D-dimer

Predictive Value of serum biomarkers: Receiver operating characteristic (ROC) curve was used to evaluate the predictive value of serum biomarkers with significant rise in patients with worse outcome (Figure 3-2).

For D-dimer, The sensitivity and specificity of the test at cut off value of D-dimer= 0.56mg/ml was 81% and 94%, respectively at $P < 0.001$.

For LDH, the sensitivity and specificity of the test at cut off value of LDH= 285.7 U/L was 71% and 76%, respectively at $P < 0.001$.

For ferritin, the sensitivity and specificity of the test at cut off value of ferritin= 387.97 ng/ml was 86% and 84%, respectively at $P < 0.001$.

For CRP, the sensitivity and specificity of the test at cut off value of CRP= 36.3 mg/L was 91% and 84%, respectively at $P < 0.001$.

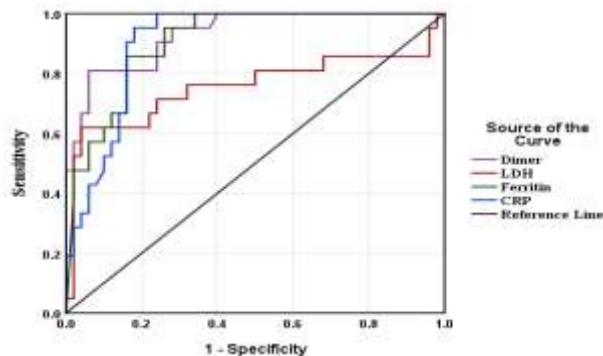


Figure 4: Receiver operating characteristic curve for D-dimer, LDH, Ferritin and CRP predicting mortality and ICU admission in patients with COVID-19

DISCUSSION

Our data revealed no significant impact ($P > 0.05$) of the gender regarding the outcome of COVID-19 disease. The literature of review showed contradictory findings regarding the influence of gender on COVID-19 outcome. For example, in China, a previous study inferred a mortality rate of 2.8 and 1.7% among men and females COVID-19 patients, respectively [24]. Another previous study conducted in China revealed a low mortality rate in women (0.4%) compared to that in men (7.2%) COVID-19 patients [25]. A previous meta-analysis study carried out on 5057 COVID-19 patients, revealed the high mortality rate among the hospitalized male group (26). In the meta-analysis study of Biswas and co-workers conducted on 64,676 coronavirus cases, it was revealed that COVID-19 male patients were significantly correlated with elevated risk of mortality compared to COVID-19 female patients ($P < 0.00001$) (27). Consistent to our data, Al-Bari, et al. inferred no significant difference between the death rate among male and females COVID-19 patients in Bangladesh (28).

Our data revealed that elderly COVID-19 aged of 59 years were the most affected patients. Similarly, a previous study conducted in Iraq showed that the highest death rate of COVID-19 patients was among elderly category of 60-69 years (29). Likewise, the study of Saeed et al., conferred that the most affected group of COVID-19 patients aged from 30-39 years (30). The meta-analysis of Nasiri conducted with 5057 patients revealed that the most affected COVID-19 patients aged 49 years (3). In Bangladesh, the death rate in the total infected COVID-19 cases was 50.2% among elderly patients of over 60-years (28). The meta-analysis study of Biswas et al., demonstrated that COVID-19 patients with age ≥ 50 years had a significant escalated risk of mortality compared to cases < 50 years (4). A previous study showed that COVID-19 patients aged ≥ 60 years demonstrated disease severity and long-term disease course compared to cases < 60 years (31). In China, the case-fatality profile among COVID-19 patients of different ages was in the following order: people > 80 years > 70 -79 years > 60 -69 years > 50 -59 years $> < 50$ years (32). Likewise in Italy, the

highest case-fatality rate was observed among COVID-19 patients of 81-90 years (33). In England and Wales the highest deaths of COVID-19 patients were among patients aged ≥ 65 years (34).

The current study reveals a significant correlation ($P < 0.05$) between the duration time and the clinical presentations of COVID-19 disease. A previous study stated that the real time -PCR test positivity and duration of the symptom duration associated significantly with initial viral load (35). The elevation of viral load is very possibly to impose overburden to body's immune response accompanied by severe illness (36). The study conducted by Wenyu et al., revealed negative significant correlation between the viral load of COVID-19 and the lymphocyte count and distinct positive correlation between the viral load and the neutrophil count and the CRP. Obviously, the correlation between the viral load of COVID-19 and the illness severity is still not fully understood. This would oblige the indispensable need to conduct further studies for further exploration and unveiling the molecular mechanisms behind this confounder.

Previous research revealed that the CRP level is positively correlated with the diameter of lesions existed in the lung with likelihood of reflection of illness severity among COVID-19 patients (37). It was reported that the elevation of neutrophil, CRP, D-dimer, and LDH might point out the progression of COVID-19 and the reduction in lymphocyte count (23). Additionally, Wenyu et al. discovered a positive linear correlation between the high viral load and the high CRP level (36).

Reportedly, D-dimer elevates in COVID-19 cases, associates with illness severity, and is trustworthy prognostic marker for mortality rate. Our finding revealed a significant difference ($P < 0.001$) among the two stratified enrolled groups in the cohort study: well discharged group and ICU admitted /died group. Likewise, Yao et al., inferred that average D-dimer level in non-survivors was significantly higher than in survivors (38). Other previous study conferred that D-dimer $> 1 \mu\text{g/ml}$ is considered one of factors imposing risks for the death among adult COVID-19 inpatients on admission (39). Yu et al., concluded that acute cases of COVID-19 display a higher level of D-dimer than those with non-acute clinical presentations, and D-dimer $>$ than $0.5 \mu\text{g/ml}$ is correlated with acute COVID-19 infection (40). A recent meta-analysis and meta-regression study has revealed that D-dimer could forecast acute and fatal consequences in patients with COVID-19 with mild sensitivity and specificity, and diagnose venous thromboembolism (VTE) with high sensitivity but low specificity (41). Conversely, previous studies revealed coagulopathy and D-dimer elevations in 3.75–68.0% of the COVID-19 inpatients and outpatients (42,43). Reportedly, the elevation of D-Dimer is restricted not only to the viral infection like COVID-19 but also to any pathologic or non-pathologic process that would raise the production of fibrin (44). Moreover, some studies verified the non-usage of D-Dimer in the context of biomarker for viral pneumonia (45). A retrospective study conducted by Yao et al, defined cut off value $> 2.14 \text{ mg/ml}$ of D-dimer upon admission forecasting mortality in inpatients with 88.2% and 71.3% of sensitivity and specificity, respectively (38).

Our finding revealed a significant increase in the level of LDH in the ICU admitted /died group compared to the well discharged group. LDH rise in COVID-19 patients evidences injuries in lung and tissue and is associated with poor diagnosis at a level $> 250 \text{ U/L}$ (46). Some studies demonstrating that LDH is not correlated with poor prognosis (46). However, a systematic review and meta-analysis study including 10399 from 21 studies conferred the association of LDH with poor diagnosis in COVID-19 patients (47). Our finding showed a positive significant correlation between the elevation of LDH and the age. However, Martha et al., revealed no association between the elevation of LDH and the age (47). The heterogeneity could be attributed to various cut-off points, lab references and tools employed in diagnosis.

Our finding revealed that the level of ferritin is significantly higher in ICU admitted/died COVID-19 patients when compared to

the well discharged group. The magnitude of inflammation existed at admission of COVID-19 patients, demonstrated by high levels of ferritin, is autonomously predictive of mortality in COVID19 inpatients (48). Vargas-Vargas and Cortés-Rojocited (49) and Bianchini et al(50) recommended the usage of the serum ferritin as one of the indicators of death in patients with COVID-19.

The association between IL-6 and iron metabolism is thoroughly known (51), however, iron parameters are not yet regarded a key biomarker to examine septic evolution. In majority of recent standards for COVID-19 (52) or in studies concerning sepsis (53), there is no reference of ferritin or other iron parameters. Whilst, in a late prospective study (54), iron parameters like ferritin and transferrin saturation levels have correlated significantly ($P=0.043$ and 0.034 , respectively) with SOFA score (Sequential Organ Failure Assessment).

In COVID19 hospitalized inpatients for acute abdomen or other surgical pathology, the acute inflammatory process has correlated with the key parameters but not with ferritin alteration. Conversely, in COVID patients, iron alteration looks to happen instantly (55). A recent meta-analysis study (56) showed that ferritin were regarded in 4 out of 16 studies analyzed, nonetheless, ferritin levels could be used in stratification of COVID19 illness's severity. This indicates the reality that a very little bit studies regard iron metabolism in COVID and non-COVID patients at the time being.

Our study considered no significant difference in IL-6 levels between the two stratified enrolled groups: well discharged group and the ICU admitted /died group. The concentration of IL-6 >24 pg/mL at beginning of evaluation did predict the progress of hypoxemia necessitating hospitalization with satisfactory sensitivity and specificity. IL-6 looks as a crucial predictor for the progress of the acute Covid-19 and could assist for primary identification of patients who are in a great need of hospitalization (57). A vital attribute of IL-6 up-regulation in Covid19 is that it leads the progress of severe lung injury implicating its usage as an initial biomarker of acute disease (58). Predominant hypothesis or speculation is that overexpression of IL-6 would play a key role in the stimulation and development of the alleged cytokine storm resulting in lung injury(59). It is assumed that IL-6 does increase the lung permeability of lung capillaries pushing the ARDS progress and furthermore does stimulate the pathway of coagulation imposing microthrombi in lung circulation and elevates the probability of risk of thrombotic event occurrence (60). The direct role of IL-6 in Covid-19 pathogenesis is verified by discoveries that IL-6 inhibition does improve the prognosis of acute Covid-19 (60,61).

Our finding revealed that there is no significant difference in the level of TGF- β among the two enrolled groups in this study: discharged well group and ICU admitted /died group. A recent study conducted by Ferreira-Gomes would suggest that in acute COVID-19 cases, SARS-CoV-2 would trigger a chronic immune reaction; instructed by TGF- β , and is distracted from itself (62). TGF- β , transforming growth factor - β , is also recognized as an outstanding immune regulator (63), and it does promote fibrosis (64), a comorbidity of acute COVID-19 cases (65). Hence, therapeutic targeting of TGF- β could be a solution to improve acute COVID-19, particularly, when putting into consideration the fibrosis-inducing capacity of TGF- β (66). The study of Shen et al., developed a practical model to identify the characteristics of cytokines storm, a signature remark in febrile and covid-19 diseases (67). This model is mainly appropriate for recognizing febrile and infectious diseases such as COVID-19. According to this model, characteristics of cytokine storm and pathogenesis of COVID-19 have been postulated to be a result of the disequilibrium in the cytokine network that occur as a consequence of the elevated biological activity of TGF- β . This would definitely impose distinct clinical presentations in the form of fever, fatigue, pneumonia, dry cough, and missing of olfactory in some cases. Research and clarification of the pathogenesis of COVID-19 will share to precision treatment.

CONCLUSIONS

In a sample of Iraqi COVID-19 patients ($n=100$), elderly people (≥ 59 years) was more susceptible for poor prognosis. Gender did not impose any significant consequence on disease outcome. The four serum biomarkers D-dimer, LDH, CRP, and ferritin might be used as indicative for poor COVID-19 disease outcome as they exhibited significant higher levels among required ICU admitted/died patients compared to their levels in the well-discharged patients. For further unveiling the molecular mechanisms behind the role of the aforementioned biomarkers in COVID-19 disease outcome, a large retrospective study would be recommended to be conducted in Iraq from the date of Coronavirus emergence till now.

REFERENCES

- 1 Samanthi . Difference Between TATA and CAAT Box June 25, 2020 Aug; 10(8): 560–574
- 2 Sophie Susen ,Charles Ambrose Tacquard³, Alexandre Godon⁴, Prevention of thrombotic risk in hospitalized patients with COVID-19 and hemostasis monitoring Critical Care volume 24, Article number: 364 (2020),
- 3 M Sakka ¹, J M Connors ², G Hékimian ³ et al Association between D-Dimer levels and mortality in patients with coronavirus disease 2019 (COVID-19): a systematic review and pooled analysis . J Med Vasc 2020 Sep;45(5):268-274.
- 4 M. Sakka,¹J.M. Connors,²G. Hékimian,³ et al Association between D-Dimer levels and mortality in patients with coronavirus disease 2019 (COVID-19): a systematic review and pooled analysis Cao X. COVID-19: immunopathology and its implications for therapy. Nat Rev Immunol. 2020;20:269–270.
- 5 Siritwan Ongchai ¹, Oraphan Somnoo ¹, Patiwat Kongdang . TGF- β 1 upregulates the expression of hyaluronan synthase 2 and hyaluronan synthesis in culture models of equine articular chondrocytes J Vet Sci . 2018 Nov 30;19(6):735-743.
- 6 M. S. Wilson and T. A. Wynn, "Pulmonary fibrosis: pathogenesis, etiology and regulation," Mucosal Immunology, vol. 2, no. 2, pp. 103–121, 2009.
- 7 Chandini Rangaswamy¹, Reiner K. Mailer Hanna Englert¹, Sandra Konrath¹Thomas Renné¹ et al .The contact system in liver injury Seminars in Immunopathology volume 43, pages 507–517 (2021)
- 8 Martijn Nolte ¹, Coert Margadant .Controlling Immunity and Inflammation through Integrin-Dependent Regulation of TGF- β Trends Cell Biol . 2020 Jan;30(1):49-59.
- 9 Misra DP, Agarwal V, Gasparian AY, Zimba O. Rheumatologists' perspective on coronavirus disease 19 (COVID-19) and potential therapeutic targets. Clin Rheumatol. 2020 Jul; 39(7):2055-2062.
- 10 Rahimmanesh, I.; Kouhpayeh, S.; Khanmahad H. The Conceptual Framework for SARS-CoV-2 Related Lymphopenia. Prepr - not peer-reviewed. 2020;(April):1–29.
- 11 Tanase DM, Gosav EM, Radu S, Ouatu A, Rezus C, Ciocoiu M, Costea CF, Floria M Arterial Hypertension and Interleukins: Potential Therapeutic Target or Future Diagnostic Marker? Int J Hypertens. 2019; 2019():3159283.
- 12 Bruno Bordallo, Mozart Bellas, Arthur Fernandes Cortez Severe COVID-19: what have we learned with the immunopathogenesis? Adv Rheumatol. 2020; 60(1): 50.
- 13 Fagone P, Ciurleo R, Lombardo SD, Iacobello C, Palermo CI, Shoenfeld Y, et al. Transcriptional landscape of SARS-CoV-2 infection dismantles pathogenic pathways activated by the virus, proposes unique sex-specific differences and predicts tailored therapeutic strategies. Autoimmun Rev. 2020 Jul; 19(7):102571
- 14 Jose Gómez-Rial,^{1,2}Irene Rivero-Calle,^{1,3}Antonio Salas,^{1,4} and Federico Martínón-Torres Role of Monocytes/Macrophages in Covid-19 Pathogenesis: Implications for Therapy Infect Drug Resist. 2020; 13: 2485–2493
- 15 Frasca D., Blomberg B.B., Paganelli R. Aging, Obesity, and Inflammatory Age-Related Diseases. Front. Immunol. 2017;8:1745.
- 16 Franceschi C, Campisi J. Exacerbated innate host response to SARS-CoV in aged non-human primates. J Gerontol A Biol Sci Med Sci. 2014 Jun; 69 Suppl 1:S4-9.
- 17 Smits SL, de Lang A, van den Brand JM, Leijten LM, van IJcken WF, Eijkemans MJ, et al PLoS Pathog. 2010 Feb 5; 6(2):e1000756
- 18 Lawrence A. Potempa,¹Ibraheem M. Rajab,¹Peter C. Hart,¹Jose Bordon,² and Rafael Fernandez-Botran. Insights into the Use of C-Reactive Protein as a Diagnostic Index of Disease Severity in COVID-19 Infections. Am J Trop Med Hyg. 2020 Aug; 103(2): 561–563.
- 19 Muus C., Luecken M.D., Eraslan G., Sikkema L., Waghray A., Heimberg G., et al. Integrated analyses of single-cell atlases reveal age, gender, and smoking status associations with cell type-specific expression of mediators of SARS-CoV-2 viral entry and highlights inflammatory programs in putative target cells. Nat Med. 2021 doi: 10.1038/s41591-020-01227-z
- 20 (Jacot et al., 2020), Jacot D., Greub G., Jaton K., Opota O. Viral load of SARS-CoV-2 across patients and compared to other respiratory viruses. Microbes Infect. 2020;22:617–621
- 21 Bastard P., Rosen L.B., Zhang Q., Michailidis E., Hoffmann H.H., Zhang Y., et al. COVID Human Genetic Effort Autoantibodies against type I IFNs in patients with life-threatening COVID-19. Science. 2020;370:eabd4585.
- 22 Norbury CC, Malide D, Gibbs JS, Bennink JR, Yewdell JW. Visualizing priming of virus-specific CD8+ T cells by infected dendritic cells in vivo. Nat Immunol. 2002 Mar; 3(3):265-71.
- 23 Ahmed N. Kaftan,¹Majid K. Hussain,¹Abdulhussain A. Algenabi,¹Farah H. Naser,² and Muslim A. Enaya Predictive Value of C-reactive Protein, Lactate Dehydrogenase, Ferritin and D-dimer Levels in Diagnosing COVID-19 Patients: a Retrospective Study. Acta Inform Med. 2021 Mar; 29(1): 45–50.

- 24 Xie J, Tong Z, Guan X, Du B, Qiu H. Clinical Characteristics of Patients Who Died of Coronavirus Disease 2019 in China. *JAMA Netw Open*. 2020 Apr 1; 3(4):e205619.
- 25 Qian J, Zhao L, Ye RZ, Li XJ, Liu YL. Age-dependent gender differences of COVID-19 in mainland China: comparative study. *Clin Infect Dis*. 2020;71(9):2488–94. doi:10.1093/cid/ciaa683.
- 26 Nasiri MJ, Haddadi S, Tahvildari A, Farsi Y, Arbabi M, Hasanzadeh S, et al. (2020) COVID-19 Clinical Characteristics, and Sex-Specific Risk of Mortality: Systematic Review and Meta-Analysis. *Front. Med*. 7:459. doi: 10.3389/fmed.2020.00459
- 27 Mohitosh Biswasa Shawonur Rahamana Tapash Kumar Biswasb Zahirul Haquec Baharudin Ibrahimd. Association of Sex, Age, and Comorbidities with Mortality in COVID-19 Patients: A Systematic Review and Meta-Analysis. *Intervirolgy* 2021;64:36–47.
- 28 Al-Bari MAA, Hossain S, Zahan MK. Exploration of sex-specific and age-dependent COVID-19 fatality rate in Bangladesh population. *World J Radiol*. 2021;13(1):1-18. doi:10.4329/wjr.v13.i1.1.
- 29 Faris Lami, Hiba Abdulrahman Rashak, Hanan Abdulghafoor Khaleel, Sinan Ghazi Mahdi, Firas Adnan, Yousef S Khader, et al. Iraq experience in handling the COVID-19 pandemic: implications of public health challenges and lessons learned for future epidemic preparedness planning. *Journal of Public Health, Volume 43, Issue Supplement_3, 2021, Pages iii19–iii28*.
- 30 Saeed BQ, Al-Shahrabi R, Bolarinwa OA (2021) Socio-demographic correlate of knowledge and practice toward COVID-19 among people living in Mosul-Iraq: A cross-sectional study. *PLoS ONE* 16(3): e0249310.
- 31 Liu Y, Mao B, Liang S, et al. Association between age and clinical characteristics and outcomes of COVID-19. *Eur Respir J* 2020; 55: 2001112 [https://doi.org/10.1183/13993003.01112-2020].
- 32 Li Q, Guan X, Wu P, Wang X, Cowling B, et al. (2020) Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. *N Engl J Med*. 33 https://www.cnbc.com/2020/03/27/why-coronavirus-deaths-are-higher-in-italy-spain-than-in-china.html.
- 34 https://www.ons.gov.uk/peoplepopulationandcommunity/birthsdeathsandmarriages/deaths/articles/coronavirusrelateddeathsbyethnicgroupenglandandwales/2march2020to10april2020#ethnic-breakdown-of-deaths-by-age-and-sex
- 35 Caplan A, Bates KW, Brioni C, Santos A, Sabatini LM, Kaul KL, et al. (2021) Clinical characteristics and viral load dynamics of COVID-19 in a mildly or moderately symptomatic outpatient sample. *PLoS ONE* 16(10): e0258970. https://doi.org/10.1371/journal.pone.0258970.
- 36 Wenyu Chen, Qinfeng Xiao, Zhixian Fang, Xiaodong Lv, Ming Yao, Min Deng. "Correlation Analysis between the Viral Load and the Progression of COVID-19", *Computational and Mathematical Methods in Medicine*, vol. 2021. https://doi.org/10.1155/2021/9926249
- 37 L. Wang, "C-reactive protein levels in the early stage of COVID-19," *Médecine et Maladies Infectieuses*, vol. 50, no. 4, pp. 332–334, 2020.
- 38 Yao, Y, Cao, J, Wang, Q, et al. D-dimer as a biomarker for disease severity and mortality in COVID-19 patients: a case control study. *J Intensive Care*. 2020;8:49.
- 39 Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet*. 2020; 395(10229):1054-62
- 40 Yu HH, Qin C, Chen M, Wang W, Tian DS. D-dimer level is associated with the severity of COVID-19. *Thromb Res*. 2020;195:219-225. doi:10.1016/j.thromres.2020.07.047
- 41 Haoting Zhan , Haizhen Chen, Chenxi Liu, Linlin Cheng, Songxin Yan, Haolong Li, Yongzhe Li. Diagnostic Value of D-Dimer in COVID-19: A Meta-Analysis and Meta-Regression. *Clinical and Applied Thrombosis/Hemostasis*. 2021. Volume 27: 1-10.
- 42 Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020;395(10223):507–13.
- 43 Wu J, Liu J, Zhao X, Liu C, Wang W, Wang D, Xu W, Zhang C, Yu J, Jiang B, et al. Clinical Characteristics of Imported Cases of COVID-19 in Jiangsu Province: A Multicenter Descriptive Study. *Clin Infect Dis*. 2020;29:ciaa199.
- 44 Linkins LA, Takach Lapner S. Review of D-dimer testing: good, bad, and ugly. *Int J Lab Hematol*. 2017;39(1):98–103.
- 45 Querol-Ribelles JM, Tenias JM, Grau E, Querol-Borras JM, Climent JL, Gomez E, Martinez I. Plasma d-dimer levels correlate with outcomes in patients with community-acquired pneumonia. *Chest*. 2004;126(4):1087–92
- 46 Li X, Xu S, Yu M, et al. Risk factors for severity and mortality in adult COVID-19 inpatients in Wuhan. *J Allergy Clin Immunol* 2020; 146:110–8.
- 47 Martha JW, Wibowo A, Pranata R. Prognostic value of elevated lactate dehydrogenase in patients with COVID-19: a systematic review and meta-analysis. *Postgraduate Medical Journal* Published Online First: 15 January 2021. doi: 10.1136/postgradmedj-2020-139542
- 48 Katia Lino, Gabriel Macedo Costa Guimarães, Lilian Santos Alves, Any Caroline Oliveira , Renan Faustino, Cintia Souza Fernandes, et al. Serum ferritin at admission in hospitalized COVID-19 patients as a predictor of mortality. *Braz. J. Infect. Dis* (2 0 2 1);25(2):101569.
- 49 Vargas-Vargas M, Cortés-Rojo C. Ferritin levels and COVID-19. *Rev Panam Salud Publica*. 2020;44:e72.
- 50 Banchini, F., Cattaneo, G.M. & Capelli, P. Serum ferritin levels in inflammation: a retrospective comparative analysis between COVID-19 and emergency surgical non-COVID-19 patients. *World J Emerg Surg* 16, 9 (2021).
- 51 Narazaki M, Kishimoto T. The two-faced cytokine IL-6 in host defense and diseases. *Int J Mol Sci*. 2018;19(11):3528.
- 52 Alhazzani W, Møller MH, Arabi YM, Loeb M, Gong MN, Fan E, et al. Surviving sepsis campaign: guidelines on the management of critically ill adults with Coronavirus Disease 2019 (COVID-19). *Intensive Care Med*. 2020;46(5):854–87.
- 53 Bateman RM, Sharpe MD, Jagger JE, Ellis CG, Solé-Violán J, López-Rodríguez M, et al. 36th International Symposium on Intensive Care and Emergency Medicine: Brussels, Belgium. 15-18 March 2016. *Crit Care*. 2016;20(Suppl 2):94
- 54 Brandtner A, Tymoszuk P, Nairz M, Lehner GF, Fritsche G, Vales A, et al. Linkage of alterations in systemic iron homeostasis to patients' outcome in sepsis: a prospective study. *J Intensive Care*. 2020;8:76.
- 55 Huang C, Wang Y., Li X. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395(10223):497–506. doi:10.1016/s0140-6736(20)30183-5.
- 56 -Zeng F, Huang Y, Guo Y, Yin M, Chen X, Xiao L, Deng G. Association of inflammatory markers with the severity of COVID-19: a meta-analysis. *Int J Infect Dis*. 2020;96:467–74.
- 57 Sabaka, P., Koščálová, A., Straka, I. et al. Role of interleukin 6 as a predictive factor for a severe course of Covid-19: retrospective data analysis of patients from a long-term care facility during Covid-19 outbreak. *BMC Infect Dis* 21, 308 (2021). https://doi.org/10.1186/s12879-021-05945-8
- 58 Aziz M, Fatima R, Assaly R. Elevated interleukin-6 and severe COVID-19: A 75-Magro G. SARS-CoV-2 and COVID-19: is interleukin-6 (IL-6) the 'culprit lesion' of ARDS onset? What is there besides Tocilizumab? *SGP130Fc. Cytokine X*. 2020;2(2):100029.
- 59 Hunter CA, Jones SA. IL-6 as a keystone cytokine in health and disease. *Nat Immunol*. 2015;16:448–57.
- 60 Giamarellou-Bourboulis EJ, Netea MG, Rovina N, et al. Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure. *Cell Host Microbe*. 2020;27(6):992–1000.e3.
- 61 Guaraldi G, Meschiarri M, Cozzi-Lepri A, et al. Tocilizumab in patients with severe COVID-19: a retrospective cohort study. *Lancet Rheumatol*. 2020;2(8):e474–84.
- 62 Ferreira-Gomes, M., Kruglov, A., Durek, P. et al. SARS-CoV-2 in severe COVID-19 induces a TGF-β-dominated chronic immune response that does not target itself. *Nat Commun* 12, 1961 (2021). https://doi.org/10.1038/s41467-021-22210-3.
- 63 Beller, A. et al. Specific microbiota enhances intestinal IgA levels by inducing TGF-beta in T follicular helper cells of Peyer's patches in mice. *Eur. J. Immunol*. 50, 783–794 (2020).
- 64 Lee, C. G. et al. Interleukin-13 induces tissue fibrosis by selectively stimulating and activating transforming growth factor beta(1). *J. Exp. Med*. 194, 809–821 (2001).
- 65 Leeming, D. J. et al. Can biomarkers of extracellular matrix remodelling and wound healing be used to identify high risk patients infected with SARS-CoV-2?: lessons learned from pulmonary fibrosis. *Respir. Res*. 22, 38 (2021).
- 66 Polak, S. B., Van Gool, I. C., Cohen, D., von der Thüsen, J. H. & van Paassen, J. A systematic review of pathological findings in COVID-19: a pathophysiological timeline and possible mechanisms of disease progression. *Mod. Pathol*. 33, 2128–2138 (2020).
- 67 Shen W-X, Luo R-C, Wang J-Q and Chen Z-S (2021) Features of Cytokine Storm Identified by Distinguishing Clinical Manifestations in COVID-19. *Front. Public Health* 9:671788.